

*Citation for published version:*

Jevglevskis, M, Lee, GL, Nathubhai, A, James, T, Threadgill, M, Woodman, T & Lloyd, M 2016, 'A Convenient Colorimetric Assay for -Methylacyl-CoA Racemase (AMACR; P504S) and Testing Of Inhibitors', Cancer Research @ Bath 13th symposium, Bath, UK United Kingdom, 27/04/16 - 27/04/16.

*Publication date:*  
2016

*Document Version*  
Other version

[Link to publication](#)

*Publisher Rights*  
Unspecified

**University of Bath**

## **Alternative formats**

If you require this document in an alternative format, please contact:  
[openaccess@bath.ac.uk](mailto:openaccess@bath.ac.uk)

### **General rights**

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

### **Take down policy**

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.



Maksims Yevglevskis,<sup>a</sup> Guat Lee,<sup>a</sup> Amit Nathubhai,<sup>a</sup> Tony D. James,<sup>b</sup> Michael D. Threadgill,<sup>a</sup> Timothy J. Woodman<sup>a</sup> and Matthew D. Lloyd<sup>a,‡</sup>

Medicinal Chemistry, Department of Pharmacy & Pharmacology, University of Bath, Claverton Down, Bath, BA2 7AY, U.K. <sup>‡</sup>email: [M.D.Lloyd@bath.ac.uk](mailto:M.D.Lloyd@bath.ac.uk)  
Department of Chemistry, University of Bath, Claverton Down, Bath, BA2 7AY, U.K.

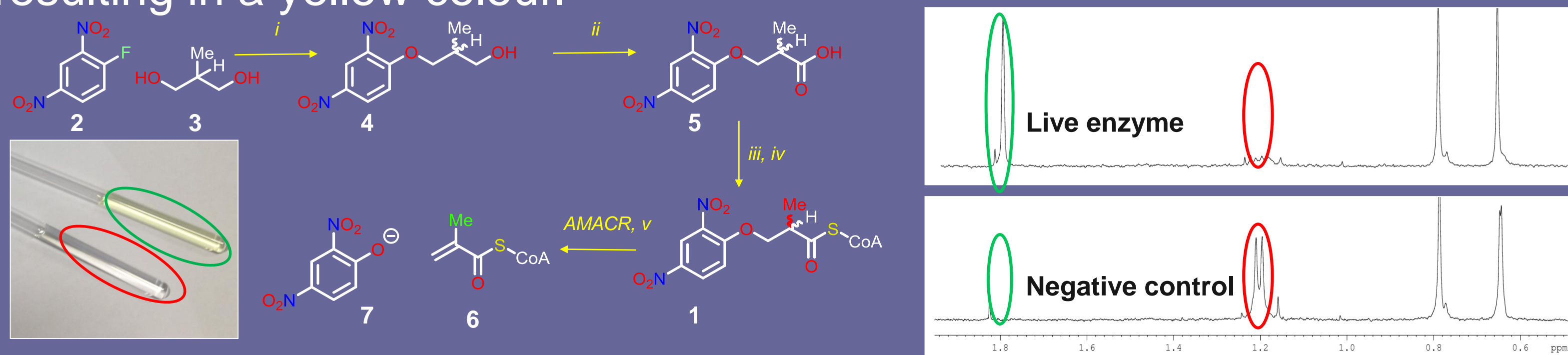
## Introduction

Branched-chain fatty acids are common in the diet and similar structures are found in medicines such as Ibuprofen and related drugs. Metabolism of branched-chain fatty acids requires that the centres bearing the methyl groups possess *S*-stereochemical configuration, but those with *R*-configuration are produced in the body and are found in the diet. Ibuprofen and related drugs require *S*-configuration for their anti-inflammatory properties, but these drugs are usually given as a mixture of *R*- and *S*-enantiomers. The enzyme  $\alpha$ -methylacyl-CoA racemase (AMACR) catalyses *R*- to *S*- conversion of 2-methylacyl-CoA derivatives of fatty acids enabling  $\beta$ -oxidation. Similarly, acyl-CoA derivatives of Ibuprofen and similar drugs are converted, resulting in pharmacological activation.<sup>1,2</sup>

AMACR levels are increased in all prostate cancers, some colon cancers and other cancers.<sup>1-3</sup> In prostate cancer, higher AMACR levels result in higher proliferation rates<sup>4</sup> and androgen-independent growth<sup>5</sup> and AMACR is recognised as a novel drug target. However, few inhibitors have been identified, largely due to the difficulties in measuring enzyme activity which makes it difficult to quantify drug potency.<sup>1</sup> AMACR catalyses the irreversible elimination of hydrogen fluoride from 3-fluoro-2-methylacyl-CoA substrates,<sup>6</sup> but translating this reaction to a convenient colorimetric or fluorometric assay has proven difficult.<sup>3</sup> 4-Nitrophenol derivatives are commonly used as colorimetric substrates for enzymes. This study reports the synthesis of a 2,4-dinitrophenol-containing AMACR substrate and the characterisation of known AMACR inhibitors using a convenient colorimetric microtitre plate assay.

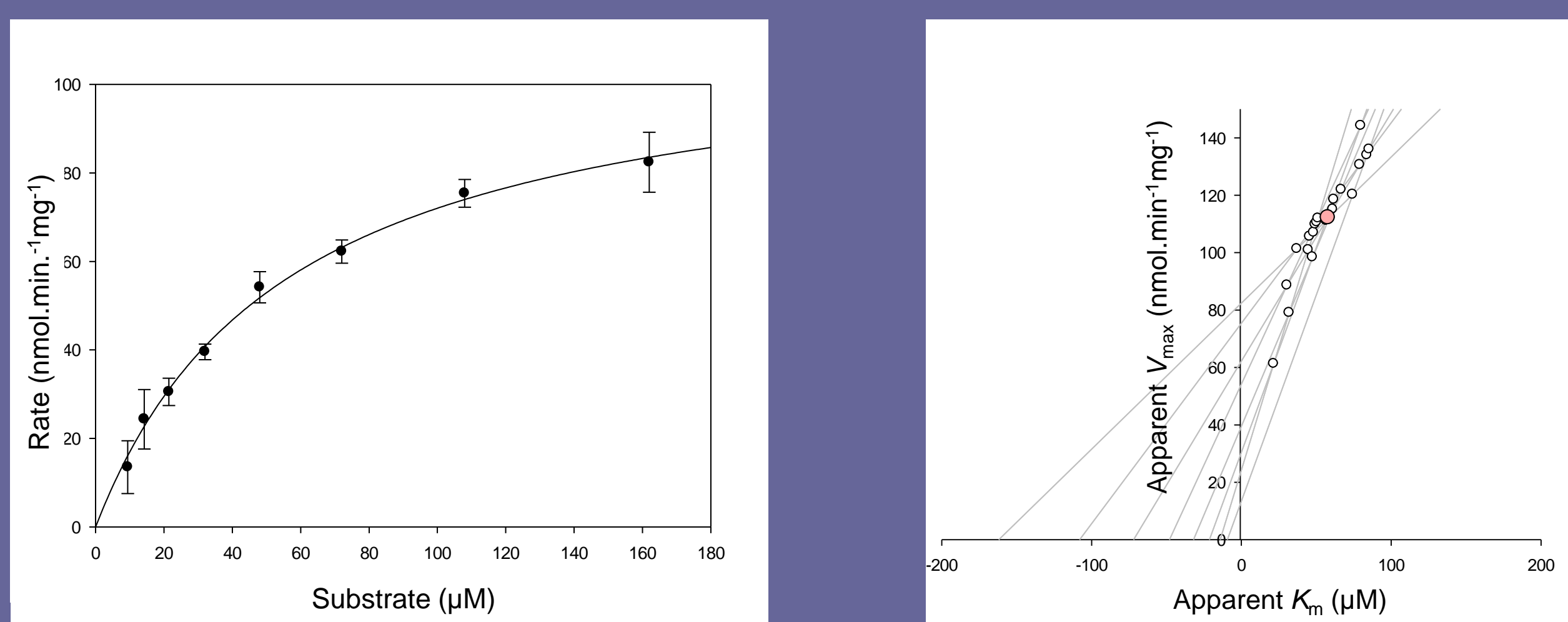
## Results and Discussion

2,4-Dinitrophenol is fully ionised at neutral pH giving a yellow colour and has a similar *pK*<sub>a</sub> to HF, which is eliminated from known AMACR substrates. Therefore an acyl-CoA derivative **1** containing 2,4-dinitrophenol was investigated. Reaction of **2** with alcohol **3** to give **4** followed by oxidation gave the racemic acid **5**, which was converted to the desired substrate **1** (Scheme 1). Incubation of **1** with recombinant human AMACR 1A resulted in formation of unsaturated product **6** and 2,4-dinitrophenol **7** resulting in a yellow colour.



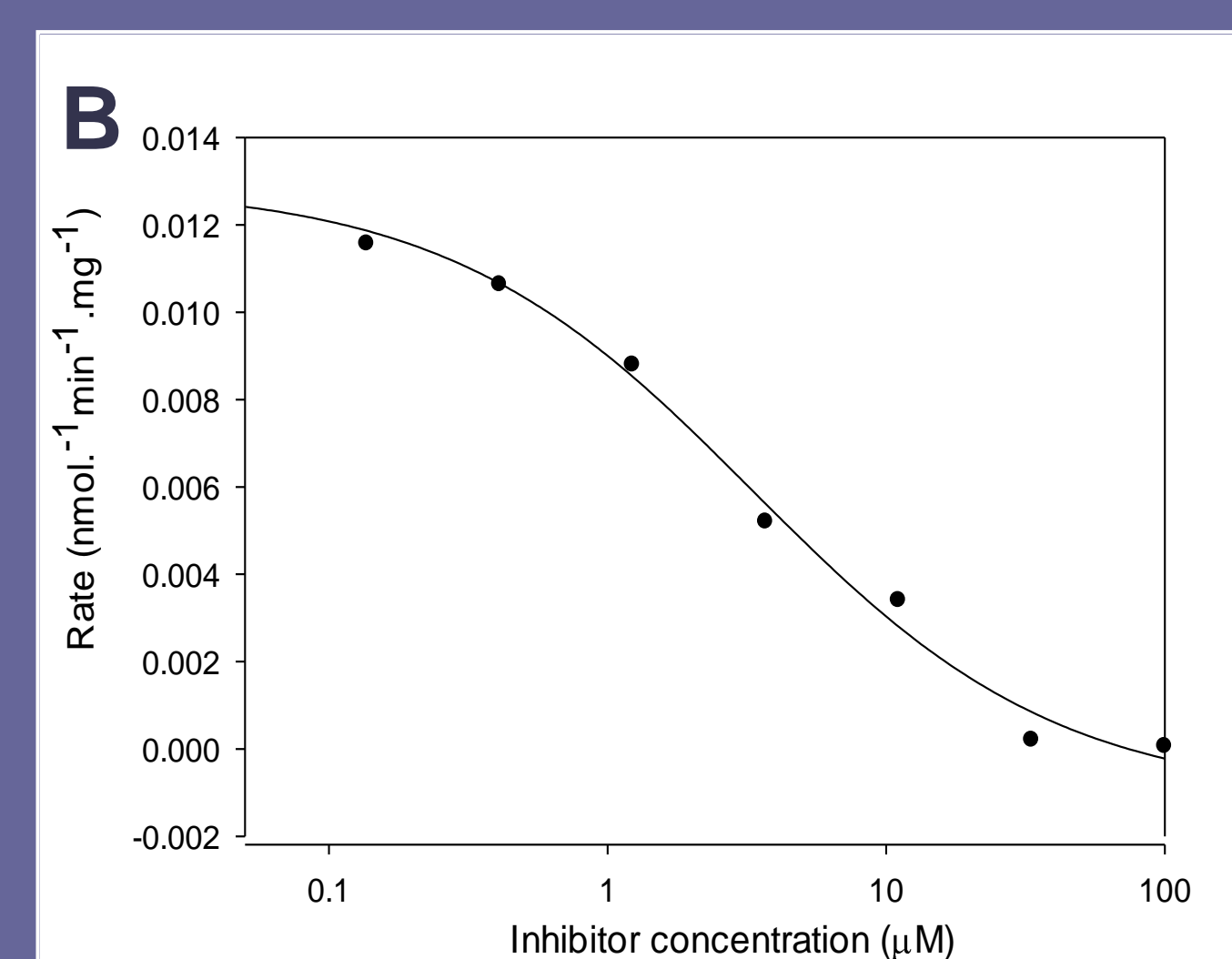
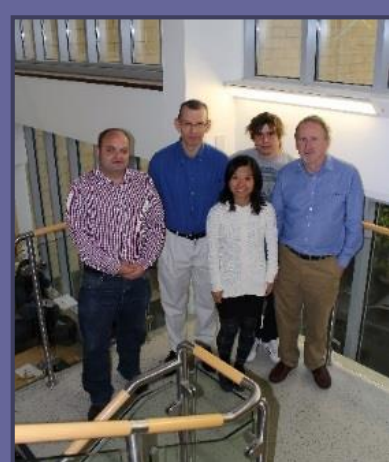
**Scheme 1:** Synthesis of novel substrate **1** and reaction with AMACR. *Reagents & conditions:* i. Na metal; ii. Jones oxidation; iii. CDI, DCM; iv. CoA-SH, NaHCO<sub>3</sub> aq./THF (1:1); v. NaH<sub>2</sub>PO<sub>4</sub>-NaOH, pH 7.4, *ca.* 77% <sup>2</sup>H<sub>2</sub>O.

AMACR was active around neutral pH and retained full activity in the presence of 8% (v/v) DMSO. Kinetic analysis of substrate **1** showed that Michaelis-Menten kinetics were observed (Figure 1), with the following parameters: *K*<sub>m</sub> = 56 ± 4.5  $\mu$ M; *V*<sub>max</sub> = 112 ± 4 nmol.min.<sup>-1</sup>mg<sup>-1</sup>; *k*<sub>cat</sub> = 0.088 s<sup>-1</sup>; *k*<sub>cat</sub>/*K*<sub>m</sub> = 1571 s<sup>-1</sup> M<sup>-1</sup>. This shows that substrate **1** is converted with ~44% of the efficiency of 3-fluoro-2-methyldecanoyl-CoA and was significantly more efficient than ‘racemisation’ of 2-methyldecanoyl-CoA (as judged by *k*<sub>cat</sub>/*K*<sub>m</sub>).<sup>6</sup>



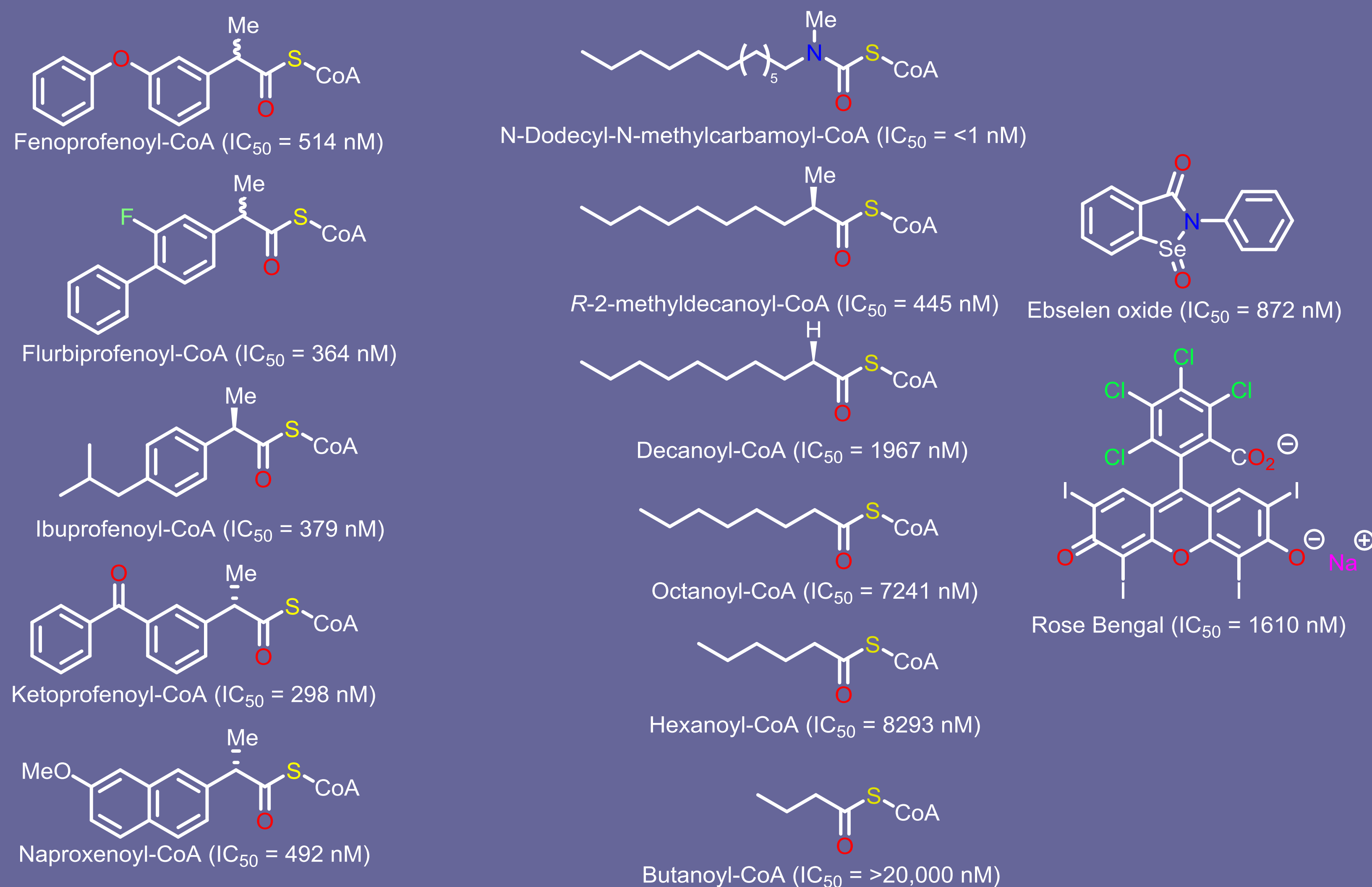
**Figure 1:** Kinetic analysis for substrate **1**.

The known inhibitor Rose Bengal<sup>7</sup> was tested to validate the method for characterisation of inhibitors (Figure 2). A dose-response curve was efficiently produced using a microtitre plate assay.



**Figure 2:** AMACR inhibition assay using Rose Bengal as an inhibitor. A. 96-Well plate showing colour change; B. Dose-response curve for Rose Bengal.

A number of other known AMACR inhibitors and substrates were tested using a dose-response curve at a fixed substrate concentration of 40  $\mu$ M. Ibuprofenoyl-CoA and related compounds are known substrates and should behave as competitive inhibitors. All of these compounds inhibited the enzyme with IC<sub>50</sub> values of *ca.* 300-500 nM. 2-Methyldecanoyl-CoA also inhibited the reaction, and was *ca.* 4x more potent than decanoyl-CoA. Inhibition was decreased in acyl-CoA esters with shorter alkyl chains. The best acyl-CoA inhibitor was *N*-dodecyl-*N*-methylcarbamoyl-CoA,<sup>8</sup> which was ~500 – 1000 x more potent than the other acyl-CoA inhibitors (as judged by IC<sub>50</sub> values). The non-specific protein modifying reagents reported by Wilson *et al.*<sup>7</sup> also inhibited the enzyme; in contrast to previous reports Ebselen behaved as a time- and concentration-dependent inactivator with a rate constant of 114 M<sup>-1</sup> s<sup>-1</sup>.



**Figure 3:** Selected acyl-CoAs and protein modifying agents shown to inhibit the conversion of substrate **1** to **6** and **7** by AMACR using the colorimetric assay.

## Conclusions

The colorimetric substrate **1** provides a convenient method for assaying AMACR and determining the behaviour and potency of inhibitors. AMACR is a promising drug target for prostate and other cancers, but until now it has been under-exploited because of the difficulties in determining enzyme activities. Inhibitors previously reported in the literature are largely limited to rationally designed acyl-CoA esters, which do not comply with Lipinski guidelines.<sup>9</sup> This new assay will facilitate the testing and development of drugs by structure-based design, rational design and lends itself to screening approaches. The latter should allow identification of inhibitors with good drug-like properties.

## Acknowledgements

This work was funded by Prostate Cancer UK (S10-03 and PG14-009), a University of Bath Overseas Research Studentship, and Shandong-Bath undergraduate exchange studentships.

## References

- M. D. Lloyd, M. Yevglevskis, G. L. Lee, P. J. Wood, M. D. Threadgill and T. J. Woodman, *Prog. Lipid Res.*, 2013, **52**, 220-230.
- M. D. Lloyd, D. J. Darley, A. S. Wierzbicki and M. D. Threadgill, *FEBS J.*, 2008, **275**, 1089-1102.
- M. Yevglevskis, G. L. Lee, J. Sun, S. Zhou, X. Sun, G. Kociok-Köhn, T. D. James, T. J. Woodman, and M. D. Lloyd, *Org. Biomol. Chem.*, DOI: 10.1039/c5ob01541c.
- S. Zha, S. Ferdinandusse, S. Denis, R. J. Wanders, C. M. Ewing, J. Luo, A. M. De Marzo and W. B. Isaacs, *Cancer Res.*, 2003, **63**, 7365-7376.
- K. Takahara, H. Azuma, T. Sakamoto, S. Kiyama, T. Inamoto, N. Ibuki, T. Nishida, H. Nomi, T. Ubai, N. Segawa and Y. Katsuoka, *Anticancer Res.*, 2009, **29**, 2497-2505.
- M. Yevglevskis, G. L. Lee, M. D. Threadgill, T. J. Woodman and M. D. Lloyd, *Chem. Commun.*, 2014, **50**, 14164-14166.
- B. A. P. Wilson, H. Wang, B. A. Nacev, R. C. Mease, J. O. Liu, M. G. Pomper and W. B. Isaacs, *Mol. Cancer Therapeut.*, 2011, **10**, 825-838.
- A. J. Carnell, R. Kirk, M. Smith, S. McKenna, L.-Y. Lian and R. Gibson, *ChemMedChem*, 2013, **8**, 1643-1647.
- C.A. Lipinski, F. Lombardo, B. W. Dominey, R. J. Feeney, *Adv. Drug Deliv. Rev.*, 2012, **64S**, 4-17.